A FOSSIL ANTHONOTHA (LEGUMINOSAE: DETARIOIDEAE: AMHERSTIEAE) SPECIES FROM THE EARLY MIOCENE (21.73 MA) OF ETHIOPIA

Aaron D. Pan,^{1,*,+} Bonnie F. Jacobs,‡ Ellen D. Currano,§ Manuel de la Estrella,∥ Patrick S. Herendeen,# and Xander M. van der Burgt**

*Museum of Texas Tech University, Texas Tech University, 3301 4th Street, Lubbock, Texas 79415, USA; †Botanical Research Institute of Texas, 1700 University Drive, Fort Worth, Texas 76107, USA; ‡Roy M. Huffington Department of Earth Sciences, Southern Methodist University, PO Box 750395, Dallas, Texas 75275, USA; §Department of Geology and Geophysics, University of Wyoming, 1000 E. University Avenue, Laramie, Wyoming 82071, USA; ∥Departamento de Botánica, Ecología y Fisiología Vegetal, Edificio C-4, Celestino Mutis, Campus de Rabanales, Universidad de Córdoba, E-14071 Córdoba, Spain; #Chicago Botanic Garden, 1000 Lake Cook Road, Glencoe, Illinois 60022, USA; and **Africa Team, Herbarium, Royal Botanic Gardens, Kew, Richmond, London TW9 3AE, United Kingdom

Editor: Kathleen B. Pigg

Premise of research. Several morphotypes of well-preserved legume leaflet compressions are known from the early Miocene Mush plant assemblages from the northwestern plateau of Ethiopia. One of these morphotypes represents a member of the Amherstieae tribe (subfamily Detarioideae) and is prevalent in the assemblage. Morphological and epidermal micromorphological characteristics are sufficient to identify it to genus. The evolutionary and biogeographic significance of the early Miocene occurrence of this genus in the uplands of Ethiopia is explored.

Methodology. Fossil laminae were compared with herbarium specimens and leaf cuticle preparations to determine their likely identity.

Pivotal results. Morphological and micromorphological characteristics support recognition of a new fossil species of *Anthonotha* (Leguminosae: Detarioideae: Amherstieae: Berlinia clade), herein named *Anthonotha shimaglae* Pan, Jacobs, Currano, Estrella, Herendeen et Burgt sp. nov. Characteristics include symmetrical, untoothed leaflets with pulvinate petiolules, short acuminate leaflet tips, eucamptodromous secondary venation, an abaxial epidermal surface possessing sinuous anticlinal cell walls, paracytic stomatal complexes, abundant trichome bases, papillate periclinal cell walls, and an adaxial epidermal surface possessing highly sinuous anticlinal cell walls. *A. shimaglae* represents the earliest-known macrofossil of the genus. The occurrence of this new species with other Detarioideae legumes, including *Englerodendron mulugetanum*, and a morphotype representing *Berlinia* or *Isoberlinia* in the early Miocene Mush plant assemblages is more analogous to high-rainfall evergreen forests of Central Africa today than East African forest types.

Conclusions. Early Miocene Mush fossils include the common occurrence of an extinct species of legume, herein named *Anthonotha shimaglae*. Prehistoric forests of the Ethiopian plateau may represent ancestral sources of currently disjunct and perhaps relictual wet forest taxa in tropical montane and coastal forests of eastern Africa today, as well as potential evolutionary sources of the Guineo-Congolian forested region.

Keywords: Africa, evolution, forest, legume, prehistoric.

Online enhancements: appendixes.

Introduction

Within the tribe Amherstieae (Leguminosae: Detarioideae), the African endemic Berlinia clade represents a diverse and ecologically important group of trees found in tropical forest and

¹ Author for correspondence; email: aaron.pan@ttu.edu.

Manuscript received January 2023; revised manuscript received March 2023; electronically published August 15, 2023.

woodland (including miombo) communities (Estrella et al. 2017, 2020; Boom et al. 2021). The fossil record indicates that this group also has had a long and substantial role in prehistoric African ecological communities throughout much of the Cenozoic, first occurring in the Paleocene and becoming ecologically prominent in the early to middle Eocene (Herendeen and Jacobs 2000; Pan et al. 2010; Romero et al. 2020).

The Berlinia clade can be subdivided into two major monophyletic branches, designated by Estrella et al. (2020) as Berlinia

International Journal of Plant Sciences, volume 184, number 7, September 2023. © 2023 The University of Chicago. All rights reserved. Published by The University of Chicago Press. https://doi.org/10.1086/725429

subclades A and B. Subclade A consists of the bambijt group (seven genera: *Brachystegia*, *Aphanocalyx*, *Michelsonia*, *Bikinia*, *Icuria*, *Julbernardia*, and *Tetraberlinia*), *Microberlinia*, *Didelotia*, and *Gilbertiodendron*. These are the dominant arborescent constituents of today's miombo communities and many African Guineo-Congolian moist and wet forest types, including almost all tropical monodominant forest formations (Bonnefille 2007; Boom et al. 2021; Pan et al. 2023). Subclade B also consists of important tropical moist and wet forest and woodland taxa, but they generally do not currently form monodominant forest formations, although the gregariousness of some species has been noted (Lebrun and Stork 2008). Berlinia subclade B includes *Anthonotha*, *Berlinia*, *Englerodendron*, *Isoberlinia*, *Librevillea*, and *Oddoniodendron* (Breteler 2010; Estrella et al. 2019, 2020; Ojeda et al. 2019).

Recently, reevaluation of the early Miocene (21.73 Ma) Mush Valley plant assemblages has provided evidence for the earliestknown structurally and taxonomically modern African tropical monodominant forest, located on the northwestern plateau of Ethiopia and representing a long-lived forest surrounding a volcanic crater lake at an elevation likely exceeding 2000 m (Faccenna et al. 2019; Currano et al. 2020; Grímsson et al. 2021; Pan et al. 2023). However, this forest differs from extant African monodominant forest formations in being dominated by an extinct species of Englerodendron, Englerodendron mulugetanum-a constituent of Berlinia subclade B (Estrella et al. 2020; Pan et al. 2023). Another morphotype in this fossil assemblage represents the second-most abundant taxon (comprising 5%-10% of the fossil leaf material in each stratigraphic level) in the assemblages and also belongs in the Amherstieae (Bush et al. 2017; Currano et al. 2020; Currano and Jacobs 2021). This morphotype, originally designated as short-drip tip to categorize it for paleoecological and paleoenvironmental studies (Currano et al. 2020; Currano and Jacobs 2021), has now been identified and represents the (currently) Central and West African wet and moist tropical forest genus Anthonotha, another member of subclade B. Today, both Anthonotha and Englerodendron coexist only in Guineo-Congolian moist and wet forest formations (Hawthorne and Jongkind 2006). Here, the new fossil species is described and its context and significance are explored.

Material and Methods

The Mush Valley plant assemblages are located on the northwestern plateau of Ethiopia between the towns of Debre Birhan and Debre Sina (Pan et al. 2012, 2014, 2023; Bush et al. 2017; Currano et al. 2020). Fossil plant compressions (leaf, wood, fruit, seeds, and pollen), along with vertebrates and insect compressions, are preserved in layers of lacustrine carbonaceous shale interbedded with thin layers of volcanic ash, most of which are less than 3 cm in thickness (Pan et al. 2012, 2014, 2023; Engel et al. 2015; Currano et al. 2020, 2021). The age of the fossils can be narrowed to ca. 21.73 Ma based on 206 Pb/ 238 U radiometric dates derived from zircons within two ashes, one immediately underlying the fossiliferous shales and the other ash occurring 5 m below that. Zircons from these ashes yield ages of 21.733 \pm 0.060 and 21.736 \pm 0.015 Ma, respectively (Tesfamichael et al. 2017).

Plant compression fossils from multiple carbonaceous shale layers were collected for paleoecological, paleoclimate, systematic, taxonomic, and floristic studies. To document and describe micromorphological characteristics of the fossil material, small (approximately 0.5-cm²) cuticle samples were removed from the fossil leaves. Fossil specimens are permanently housed at the National Museum of Ethiopia, Addis Ababa. Prepared fossil cuticle microscope slides and modern leaf cuticle microscope slides from herbarium comparative samples are housed in the Roy M. Huffington Department of Earth Sciences at Southern Methodist University, Dallas, Texas and the Chicago Botanic Garden, Glencoe, Illinois, respectively (app. A1; Pan et al. 2012, 2014, 2023; Currano et al. 2020).

Fossil specimens were compared to extant Leguminosae specimens from herbaria at the United States National Museum of Natural History (US), Missouri Botanical Garden (MO), and Botanical Research Institute of Texas (BRIT) and from highresolution digital images from the Muséum national d'Histoire naturelle (MNHN) and Global Plants on JSTOR (app. A2). Supplementary comparisons were based on published material from floras and guides (Brenan 1971; Hawthorne and Jongkind 2006; Breteler 2010). Preparation methods for herbarium and fossil leaflet cuticle and venation are provided in Herendeen and Dilcher (1990) and Pan et al. (2014), respectively.

Results

Leguminosae Jussieu 1789

Detarioideae Burmeist. 1837

Amherstieae Benth. (Hooker) 1840

Anthonotha P. Beauv. 1806

Anthonotha shimaglae Pan, Jacobs, Currano, Estrella, Herendeen et Burgt sp. nov.

Etymology. The species name, shimaglae (ሽማግሌ), means "elder" in Amharic and references this species as representing the oldest-known macrofossil record of the genus.

Diagnosis. Symmetrical, elliptic to obovate untoothed (entiremargined) microphyllous to mesophyllous leaflets with terete pulvinate petiolules and short acuminate leaflet tips. Basal leaflets with reduced length-to-width ratios compared to apical leaflets. Canaliculate leaflet midribs. Secondary venation eucamptodromous. Tertiary venation sinuous and opposite percurrent. Abaxial anticlinal cell walls sinuous. Stomata paracytic. Abaxial trichome bases present and abundant. Papillate abaxial periclinal cell walls. Highly sinuous adaxial anticlinal cell walls.

Description. Symmetrical unlobed, untoothed (entiremargined), elliptic to obovate leaflets with terete pulvinate short petiolules (figs. 1, 2). Microphyllous to mesophyllous leaflet lamina generally range in size from ca. 5–17 cm in length and 2.7–7 cm in width, typically with length-to-width ratios ranging from 1.6:1 to 2.6:1. Leaflet midribs are canaliculate. The leaflet apex is obtuse with a short acuminate drip tip. The leaflet base is convex to rounded in shape, is obtuse in angle, and ranges from symmetrical to slightly asymmetrical. Primary venation is pinnate. Secondary venation is eucamptodromous. The secondary veins are regularly spaced, except for three crowded pairs at the base; have excurrent attachment; and generally possess uniform acute angles, except those at the base, which are high angled.



Fig. 1 Fossil and extant Anthonotha apical leaflets. A, Anthonotha shimaglae Pan, Jacobs, Currano, Estrella, Herendeen et Burgt sp. nov. (holotype; MU41-54). B, Anthonotha shimaglae (paratype; MU41-44A #3). C, Anthonotha stipulacea (MNHN-P-P03503989). Scale bar = 10 mm.

Leaflets typically possess (10–)11–14(–15) pairs of secondary veins (fig. 1). Intersecondary veins are present in most intercostal areas, with proximal courses parallel to major secondaries, reticulating distal courses, and lengths greater than 50% of the length of the subjacent secondary. Tertiary venation is sinuous and generally opposite percurrent. Epimedial tertiary veins have admedial courses obtuse to the midvein and exmedial course parallel to intercostal tertiary veins. Quaternary and quintenary venation is irregular reticulate, and the areolation is moderately developed.

Cuticle. Stomatal complexes are paracytic with one subsidiary cell typically larger than the other (fig. 3*A*). Stomata are prominent on the abaxial surface and are rare (or absent) on the adaxial surface (fig. 3*A*, 3*B*). Subsidiary cells range in size from 15.2 to 22.4 μ m in length (fig. 3*A*). The anticlinal cell walls on the abaxial surface are sinuous, and those on the adaxial leaf surface are highly sinuous (fig. 3*A*, 3*B*). The abaxial epidermal cells are generally rectangular in shape and are typically 12–24 μ m in length and 4.8–23.2 μ m in width (fig. 3*A*; app. A3). Papillae are present, abundant, and centrally located on abaxial periclinal cell surfaces (fig. 3*A*; app. A3). Hairs (trichomes) are abundant on the abaxial surface, based on the presence of thickened trichome bases (fig. 3*A*; app. A3). Hair bases are surrounded by radial basal cells (fig. 3*A*; app. A3).

Holotype. MU41-54. National Museum of Ethiopia, Addis Ababa (fig. 1*A*).

Paratypes. MU23-34 (cuticle, fig. 3A; app. A3), MU17-40 (fig. 2A; cuticle, app. A3), MU27-45 #1 (fig. 2B), MU30-8A, MU41-44A #3 (fig. 1B; cuticle, app. A3), MU41-61 (cuticle; fig. 3B). Noted additional material. MU13-18A #4, MU17-15A (cuticle), MU18-2A #1, MU18-9, MU18-11A #4 (cuticle; app. A3), MU23-34 (app. A4), MU23-32B #1, MU23-52 #4, MU23-64, MU29-40 #2 (app. A4), MU30-8A (leaflet and cuticle; apps. A3, A4); MU30-36B (app. A4), MU31-15 #1, and MU41-3A.

Comments

Leaflet lamina characteristics of the fossil align with four genera in the Berlinia clade of the Amherstieae (Fabaceae: Detarioideae), *Anthonotha*, *Berlinia*, *Englerodendron*, and *Isoberlinia*, which all belong to Berlinia subclade B (Estrella et al. 2020; Pan et al. 2023). The entire-margined, symmetrical leaflets possessing eucamptodromous venation, acuminate apices, and terete pulvinate petiolules are particularly similar to those of *Anthonotha* and *Englerodendron*, although the absence of twisted petiolules potentially excludes *Englerodendron* (figs. 1, 2). These characters are also present on the obovate, more rounded leaflets (with length-to-width ratios of around 1.6:1 to 1.7:1; fig. 2), which likely represent basal leaflets (apical leaflets have greater length-to-width ratios), again comparable to *Anthonotha* (fig. 2C). In addition, leaflet cuticle micromorphological characteristics are particularly diagnostic and match



Fig. 2 Fossil and extant Anthonotha basal leaflets. A, Anthonotha shimaglae (paratype; MU17-40). B, Anthonotha shimaglae (paratype; MU27-45 #1). C, Anthonotha macrophylla (MNHN-P-P0085429). Scale bar = 10 mm.

exclusively with the genus Anthonotha (fig. 3). These include the presence of abundant trichome bases and periclinal papillae on abaxial leaflet surfaces of the fossil and extant species observed, including A. cladantha (fig. 3C, 3D), A. crassifolia (fig. 3E, 3F), A. gilletii (fig. 3G, 3H), A. lamprophylla (not shown), and A. macrophylla (fig. 31, 3J). These characteristics have not been observed in any other members of the Berlinia clade or other members of the Amherstieae examined in this study or in Pan et al. (2023). Anthonotha shimaglae differs from all observed extant species, excluding A. gilletii, in having sinuous anticlinal cell walls on the adaxial leaflet surface (fig. 3B, 3H). Anthonotha gilletii differs from the fossil taxon in generally having fewer secondary veins (8-11 pairs) on the leaflets (Breteler 2010). The age of the fossil morphotype, its geographic location far from any existing population of Anthonotha species, and its laminar morphological and micromorphological characteristics warrant its designation as a new species.

Discussion

Anthonotha sensu stricto is a small genus of ca. 17 species consisting mainly of shrubs and small- to medium-sized trees, although A. fragrans can become large trees up to 45 m tall and with stem diameters of 130 cm. Most of the species are found in tropical moist and wet forest formations in the Guineo-Congolian region of Africa, but there are a few exceptions: A. noldeae, isolated populations of which are found in central Angola and montane forests of western Tanzania having volcanic soils (fig. 4; Brenan 1971; Breteler 2010), and A. crassifolia, which occurs in moist tropical forests and savannah woodlands (Breteler 2010). Anthonotha macrophylla is the most widespread and common species in the genus, occurring mainly in secondary tropical rainforest communities extending from Guinea to the eastern Democratic Republic of the Congo (Breteler 2010). Although A. shimaglae lived in an upland moist or wet tropical forest on volcanic-derived soils, a habitat broadly similar to that supporting the extant A. noldeae, A. shimaglae leaflets are morphologically more similar in appearance to other species of Anthonotha, including A. brieyi, A. cladantha, A. crassifolia, A. lamprophylla, A. pellegrini, A. pynaertii, A. stipulacea (fig. 1C), and A. xanderi.

It is significant that the foliar characteristics of *Anthonotha*, particularly the tomentose leaflets of extant species, can be recognized in the fossil record and that an additional characteristic, papillate periclinal abaxial cell walls, is also constant (Brenan 1971; Pan et al. 2023; fig. 3). These leaf micromorphological characters are likely synapomorphies for the genus, and, given their long persistence, likely provide ecological/evolutionary fitness (Lieberman and Lieberman 1984; Wang et al. 2021). If their function was to hinder invertebrate foliar herbivory in the early Miocene Mush forest community, their efficacy is limited as shown by a recent study of *A. shimaglae* leaflets, over 40% of which displayed insect feeding damage (Currano and Jacobs 2021).

Anthonotha shimaglae specimens represent the earliest and only known macrofossils of the genus, although the palynological record indicates that it may have a long geologic history in Africa represented by some occurrences of the pollen morphospecies, *Striatopollis catatumbus* (Oboh et al. 2005; Romero



Fig. 3 Fossil and extant *Anthonotha* leaflet cuticle. *A*, *C*, *E*, *G*, *I*, Abaxial epidermal surface. Black arrows indicate an example of a papilla; white arrows indicate an example of a trichome base. *B*, *D*, *F*, *H*, *J*, Adaxial epidermal surface. *A*, *Anthonotha shimaglae* (paratype; MU23-34 #2). *B*, *Anthonotha shimaglae* (paratype; MU41-61). *C*, *D*, *Anthonotha cladantha* (MO3766337). *E*, *F*, *Anthonotha crassifolia* (K H440/95 69 and MO5086876). *G*, *H*, *Anthonotha gilletti* (MO3254528). *I*, *J*, *Anthonotha macrophylla* (US3299348 and MO2111556). Scale bars = $20 \mu m$.



Fig. 4 Distribution map and terrestrial ecoregion occurrence of extant *Anthonotha* (circles) and location of the fossil species, *Anthonotha shimaglae* (star). Modern distributions derived from Breteler (2010). This map was created with Simplemappr (https://www.simplemappr.net/).

et al. 2020). Romero et al. (2020) compared several records of S. catatumbus with various extant Amherstieae taxa and provided evidence that they can be differentiated taxonomically to generic affinity with reasonable certainty using airyscanning imaging in conjunction with deep learning classification methods. In the study, Romero et al. (2020) used several Amherstieae genera (and species) including two species of Anthonotha, A. leptorrhachis and A. macrophylla, when trying to identify the taxonomic affinities of S. catatumbus from African and South American palynological records. Recently, however, A. leptorrhachis has been transferred to the genus Englerodendron as Englerodendron leptorrhachis (Estrella et al. 2019). With this taxonomic change, African fossil S. catatumbus occurrences in the Romero et al. (2020) study that strongly indicate Anthonotha affinity should be considered to represent records of the Anthonotha clade (Anthonotha sensu stricto + Englerodendronsensu lato), not the genus alone (Estrella et al. 2019, 2020). With this taxonomic change, a Ypresian occurrence of S. catatumbus from the Anambra Basin, Nigeria (Oboh et al. 2005), identified as Anthonotha in Romero et al. (2020), represents the earliest record of either Anthonotha sensu stricto or Englerodendronsensu lato (Estrella et al. 2019, 2020). Striatopollis catatumbus and Anthonotha-type pollen from the Paleogene and early Miocene of northeastern Africa (Egypt, Ethiopia, South Sudan, and Sudan; Eisawi and Schrank 2008; El Afty et al. 2022; Yemane et al. 1987), while confidently being of Amherstieae affinity, have not been examined in the detail provided by, for example, Grímsson et al. (2019a, 2021), Romero et al. (2020), or Pan et al. (2023), and cannot provide a substantiated generic identification to either confirm or reject the presence of the Anthonotha or Englerodendron at these localities-although the macrofossil record of both genera at Mush indicates their presence on the Ethiopian plateau in the early Miocene. A taxonomic revision using higher resolution available from combining singlegrain light and scanning electron microscopy of these assemblages and others from the Paleogene and Miocene of tropical and northeastern Africa would provide a better understanding of the complex evolutionary history and ecology of Amherstieae in communities of these regions.

The occurrence of A. shimaglae in the early Miocene of the Ethiopian Plateau has implications for the evolutionary biogeography of the Berlinia clade and more generally for Africa's tropical vegetation. Today, the distribution of low-latitude, moist forests is characterized by a contrast between the nearly continuous block across the western and central regions (the Guineo-Congolian forests) and the isolated occurrences of submontane, montane, and coastal forests to the east of the Albertine Rift and Lake Victoria (fig. 4, green areas). Eastern Africa's forests are considered to be disjunct from the main forest block, although the phylogenetic histories of East-West relatives are complex and may involve more than one prior connection (e.g., Couvreur et al. 2008, 2021). In this regard, the six extant genera comprising Berlinia subclade B most often occur in forest communities, with the genus Isoberlinia differing from the others by its presence as a dominant and widespread component of deciduous wooded savannas of the Sudanian ecoregions (fig. 4, tan and amber regions). The extant forest-dwelling relatives of Anthonotha shimaglae repeat the greater disjunct forest pattern: few species exist outside the Guineo-Congolian region of Central and West Africa, and they become particularly rare east of the Albertine Mountains and Congo Basin. In fact, only four species of subclade B occur in the eastern section of the continent, each representing a different genus. Berlinia orientalis occurs in coastal forests of Tanzania and Mozambique on sandy soils (Clarke and Robertson 2000; Mackinder and Pennington 2011); Englerodendron usambarense and Isoberlinia scheffleri are endemic to the submontane and montane forests of the Eastern Arcs (Usambaras: *E. usambarense* and *I. scheffleri*; Udzungwas: *I. scheffleri*) in Tanzania (Brenan 1971; Schulman et al. 1998; Lovett et al. 2006). Interestingly, the sole species of *Anthonotha* found outside the Guineo-Congolian region is *A. noldeae*, which is not the most similar morphologically to the fossil (see above). Nevertheless, the co-occurrence of multiple subclade B taxa from the early Miocene Mush forest—including *Englerodendron mulugetanum*, *A. shimaglae*, and a leaf(let) morphotype referred to as legume 4 that either represents *Berlinia* or *Isoberlinia* (Bush et al. 2017; Currano et al. 2020)—can be found together today only in Guineo-Congolian rainforests, especially the high-rainfall (2000–3000 mm/yr), hygrophilous evergreen forests found in Gabon and Cameroon (Bonnefille 2007).

The forests that grew on the Ethiopian plateau during the late Oligocene and early Miocene need to be considered both as ancestral sources to currently disjunct moist and wet forest taxa isolated in the tropical montane and coastal forests of eastern Africa today and as potential evolutionary sources for the Guineo-Congolian region itself (Yemane et al. 1987; Pan et al. 2006, 2010, 2012, 2014, 2023; Pan 2007; Pan and Jacobs 2009; Bonnefille 2010; Jacobs et al. 2010; Currano et al 2011, 2020; Grímsson et al. 2019*a*, 2019*b*, 2021). As to the modern disjunct subclade B genera in East Africa, it remains to be seen whether they are relicts, the product of long-distance dispersal, or some of both.

The genera today distributed to the north and south of the main Guineo-Congolian forest block are adapted to drier climates. These include members of Berlinia subclade A (the bambijt clade [namely, *Brachystegia* and *Julbernardia*]), major components of (Zambezian) miombo woodlands in southeastern and south-central Africa. The genus *Isoberlinia* in subclade B is a major component of northern (Sudanian) woodlands on the continent, convergently filling the similar niche of subclade A members to the south. A key aspect of the diverse presence of the Berlinia clade in the prehistoric forests of Mush and, potentially, other portions of northeastern Africa during the early Miocene is that the northeastern and eastern regions of the continent may be the pivotal focal point from which to examine evolutionary divergence(s) among those taxa adapted to either wetter or drier environments. What remains to be discovered is whether the isolated species of Berlinia subclade B found in moist and wet forests in eastern Africa today represent members of Guineo-Congolian genera that reinvaded these forests during optimal environmental conditions or whether they represent the older remnants of clades that had greater ecological impacts in the late Paleogene and early Neogene, now representing shadows of their former glory.

Acknowledgments

We thank the Authority for Research and Conservation of Cultural Heritage for permitting us to conduct research in the Mush Valley, as well as the director and staff of the National Museum of Ethiopia for assisting us in our research endeavor. We are very thankful to our colleagues for their contribution to our fieldwork, including M. Clemens, D. Danehy, M. Feseha, L. Jacobs, J. Noret, and N. Tabor. We are also grateful to the herbaria, particularly BRIT, KEW, MNHM, MO, and US, and their staff. We are also very thankful for the warmth and generosity of the people of the Lower and Upper Mush Valley who made our research visits more akin to homecomings. We are also very appreciative to the anonymous two reviewers whose comments and suggestions improved this work. Funding was provided by US National Science Foundation grants EAR 1053549 (Bonnie F. Jacobs) and EAR 1052478 (Ellen D. Currano), and National Geographic Society grant NGS CRE 8816-10 (Ellen D. Currano).

Literature Cited

- Bonnefille R 2007 Rainforest responses to past climatic changes in tropical Africa. Pages 115–170 *in* MB Bush, JR Flenley, eds. Tropical rainforest responses to climatic change. Praxis, Chichester.
- Bonnefille R 2010 Cenozoic vegetation, climate changes and hominid evolution in tropical Africa. Glob Planet Change 72:390–411.
- Boom AF, J Migliore, E Kaymak, P Meerts, OJ Hardy 2021 Plastid introgression and evolution of African miombo woodlands: new insights from the plastome-based phylogeny of *Brachystegia* trees. J Biogeogr 48:933–946.
- Brenan JPM 1971 Leguminosae subfamily Caesalpinioideae. Pages 1– 230 *in* E Milne-Redhead, RM Polhill, eds. Flora of tropical East Africa. Whitefriars, London.
- Breteler FJ 2010 Revision of the African genus Anthonotha (Leguminosae, Caesalpinioideae). Plant Ecol Evol 143:70–99.
- Bush RT, J Wallace, ED Currano, BF Jacobs, FA McInerney, RE Dunn, NJ Tabor 2017 Cell anatomy and leaf δ^{13} C as proxies for shading and canopy structure in a Miocene forest from Ethiopia. Palaeogeogr Palaeoclimatol Palaeoecol 485:593–604.
- Clarke GP, SA Robertson 2000 Vegetation communities. Pages 83– 102 in ND Burgess, GP Clarke, eds. Coastal forests of eastern Africa. IUCN, Gland.
- Couvreur TLP, LW Chatrou, MSM Sosef, JE Richardson 2008 Molecular phylogenetics reveal multiple tertiary vicariance origins of the African rain forest trees. BMC Biol 6:54.

- Couvreur TLP, G Dauby, A Blach-Overgaard, V Deblauwe, S Dessein, V Droissart, OJ Hardy, et al 2021 Tectonics, climate and the diversification of the tropical African flora and fauna. Biol Rev 96:16–51.
- Currano ED, BF Jacobs 2021 Bug-bitten leaves from the early Miocene of Ethiopia elucidate the impacts of plant nutrient concentration and climate on insect herbivore communities. Glob Planet Change 207:103655.
- Currano ED, BF Jacobs, RT Bush, A Novello, M Feseha, F Grímsson, FA McInerney, et al 2020 Ecological dynamic equilibrium in an early Miocene (21.73 Ma) forest, Ethiopia. Palaeogeogr Palaeoclimatol Palaeoecol 539:109425.
- Currano ED, BF Jacobs, AD Pan 2021 Is Africa really an "odd man out"? evidence for diversity decline across the Oligocene-Miocene boundary. Int J Plant Sci 182:551–563.
- Currano ED, BF Jacobs, AD Pan, NJ Tabor 2011 Inferring ecological disturbance in the fossil record: a case study from the late Oligocene of Ethiopia. Palaeogeogr Palaeoclimatol Palaeoecol 309:242–252.
- Eisawi A, E Schrank 2008 Upper Cretaceous to Neogene palynology of the Melut Basin, southeast Sudan. Palynology 32:101–129.
- El Afty H, SY El Beialy, MK Zobaa, AA Taha, D Uhl 2022 A snapshot into the Oligocene vegetation of the Tethyan southern shores: new fossil pollen evidence from North Africa (Egypt). Palynology 46:1–17.
- Engel MS, ED Currano, BF Jacobs 2015 The first mastotermitid termite from Africa (Isoptera: Mastotermitidae): a new species of *Mastotermes* from the early Miocene of Ethiopia. J Paleontol 89:1038–1042.

- Estrella M de la, S Cervantes, SB Janssens, F Forest, OJ Hardy, DI Ojeda 2020 The impact of rainforest area reduction in the Guineo-Congolian region on the tempo of diversification and habitat shifts in the Berlinia clade (Leguminosae). J Biogeogr 47:2728–2740.
- Estrella M de la, F Forest, JJ Wieringa, M Fougère-Danezan, A Bruneau 2017 Insights on the evolutionary origin of Detarioideae, a clade of ecologically dominant tropical African trees. New Phytol 214:1722–1735.
- Estrella M de la, JJ Wieringa, FJ Breteler, DI Ojeda 2019 Re-evaluation of the genus *Englerodendron* (Leguminosae–Detarioideae) including *Isomacrolobium* and *Pseudomacrolobium*. Aust Syst Bot 32:564–571.
- Faccenna C, P Glišović, A Forte, TW Becker, E Garzanti, A Sembroni, Z Gvirtzman 2019 Role of dynamic topography in sustaining the Nile River over 30 million years. Nat Geosci 12:1012–1017.
- Grímsson F, SA Graham, M Coiro, BF Jacobs, A Xafis, FH Neumann, L Scott, J Sakala, ED Currano, R Zetter 2019*a* Origin and divergence of Afro-Indian Picrodendraceae: linking pollen morphology, dispersal modes, fossil records, molecular dating and paleogeography. Grana 58:227–275.
- Grímsson F, BF Jacobs, L Johan, CH Van Valkenburg, JJ Wieringa, A Xafis, N Tabor, AD Pan, R Zetter 2019 *Sclerosperma* fossils from the late Oligocene of Chila, north-western Ethiopia. Grana 58:81–98.
- Grímsson F, S Ulrich, M Coiro, SA Graham, BF Jacobs, ED Currano, A Zafis, R Zetter 2021 *Hagenia* from the early Miocene of Ethiopia: evidence for possible niche evolution. Ecol Evol 11:5164–5186.
- Hawthorne W, C Jongkind 2006 Woody plants of western African forests: a guide to the forest trees, shrubs and lianes from Senegal to Ghana. Kew Publishing, Kew.
- Herendeen PS, DL Dilcher 1990 Reproductive and vegetative evidence for the occurrence of *Crudia* (Leguminosae, Caesalpinioideae) in the Eocene of southeastern North America. Bot Gaz 151:402–413.
- Herendeen P, BF Jacobs 2000 Fossil legumes from the middle Eocene (46.0 Ma) Mahenge flora of Singida, Tanzania. Am J Bot 87:1358–1366.
- Jacobs BF, AD Pan, CR Scotese 2010 A review of the Cenozoic vegetation history of Africa. Pages 57–72 *in* L Werdelin, WJ Sanders, eds. Cenozoic mammals of Africa. University of California Press, Berkeley.
- Lebrun J-P, AL Stork 2008 Tropical African flowering plants: ecology and distribution. Vol 3. Mimosaceae–Fabaceae (incl. *Derris*). Èditions Conservatoire de Jardin botaniques de la Ville de Genève, Geneva.
- Lieberman D, M Leiberman 1984 The causes and consequences of synchronous flushing in a dry tropical forest. Biotropica 16:193–201.
- Lovett JC, CK Ruffo, RE Gereau, JRD Taplin 2006 Field guide to the moist forest trees of Tanzania. Society for Environmental Exploration, London.
- Mackinder BA, RT Pennington 2011 Monograph of *Berlinia* (Leguminosae). Systematic Botany Monographs 91. American Society of Plant Taxonomists, Laramie, WY.

- Oboh FE, CG Obi, CA Jaramillo 2005 Lithofacies, palynofacies, and sequence stratigraphy of Paleogene strata in southeastern Nigeria. J Afr Earth Sci 41:79–102.
- Ojeda DI, E Koenen, S Cervantes, M de la Estrella, E Banguera-Hinestroza, SB Janssens, J Migliore, BB Demenou, A Bruneau, F Forest, et al 2019 Phylogenomic analyses reveal an exceptionally high number of evolutionary shifts in a florally diverse clade of African legumes. Mol Phylogenet Evol 137:156–167.
- Pan AD 2007 The late Oligocene (28–27 Myr) Guang River flora from the northwestern plateau of Ethiopia. PhD diss. Southern Methodist University, Dallas, TX.
- Pan AD, ED Currano, BF Jacobs, M Feseha, N Tabor, PS Herendeen 2012 Fossil Newtonia (Fabaceae: Mimoseae) seeds from the early Miocene (22–21 Ma) Mush Valley in Ethiopia. Int J Plant Sci 173:290–296.
- Pan AD, BF Jacobs 2009 The earliest record of the genus Cola (Malvaceae sensu lato: Sterculioideae) from the Late Oligocene (28–27 Ma) of Ethiopia and leaf characteristics within the genus. Plant Syst Evol 283:247–262.
- Pan AD, BF Jacobs, RT Bush, M de la Estrella, F Grímsson, PS Herendeen, XM van der Burgt, ED Currano 2023 First evidence of a monodominant (*Englerodendron*, Amherstieae, Detarioideae, Leguminosae) tropical moist forest from the early Miocene (21.73 Ma) of Ethiopia. PLoS ONE 18:e0279491.
- Pan AD, BF Jacobs, ED Currano 2014 Dioscoreaceae fossils from the late Oligocene and early Miocene of Ethiopia. Bot J Linn 175:17– 28.
- Pan AD, BF Jacobs, J Dransfield, WJ Baker 2006 The fossil history of palms (Arecaceae) in Africa and new records from the Late Oligocene (28–27 Mya) of north-western Ethiopia. Bot J Linn Soc 151:69–81.
- Pan AD, BF Jacobs, PS Herendeen 2010 Detarieae sensu lato (Fabaceae) from the late Oligocene (27.23 Ma) Guang River flora of north-western Ethiopia. Bot J Linn 163:44–54.
- Romero IC, S Kong, CC Fowlkes, C Jaramillo, MA Urban, F Oboh-Ikuenobe, C D'Apolito, SW Punyasena 2020 Improving the taxonomy of fossil pollen using convolutional neural networks and superresolution microscopy. Proc Natl Acad Sci USA 117:45.
- Schulman L, L Junikka, A Mndolwa, I Rajabu 1998 Trees of Amani Nature Reserve, NE Tanzania. Helsinki University, Helsinki.
- Tesfamichael T, B Jacobs, N Tabor, L Michel, E Currano, M Feseha, R Barclay, J Kappelman, M Schmitz 2017 Settling the issue of "decoupling" between atmospheric carbon dioxide and global temperature: [CO₂]_{atm} reconstructions across the warming Paleogene-Neogene divide. Geology 45:999–1002.
- Wang X, C Shen, P Meng, G Tan, L Lv 2021 Analysis and review of trichomes in plants. BMC Plant Biol 21:70.
- Yemane K, C Robert, R Bonnefille 1987 Pollen and clay mineral assemblages of a late Miocene lacustrine sequence from the northwestern Ethiopian highlands. Palaeogeogr Palaeoclimatol Palaeoecol 60:123–141.